

# Persistent gating deficit and increased sensitivity to NMDA receptor antagonism after puberty in a new mouse model of the human 22q11.2 microdeletion syndrome: a study in male mice

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**Background:** The hemizygous 22q11.2 microdeletion is a common copy number variant in humans. The deletion confers high risk for neurodevelopmental disorders, including autism and schizophrenia. Up to 41% of deletion carriers experience psychotic symptoms. **Methods:** We present a new mouse model (Df(h22q11)/+) of the deletion syndrome (22q11.2DS) and report on, to our knowledge, the most comprehensive study undertaken to date in 22q11.2DS models. The study was conducted in male mice. **Results:** We found elevated postpubertal *N*-methyl-D-aspartate (NMDA) receptor antagonist-induced hyperlocomotion, age-independent prepulse inhibition (PPI) deficits and increased acoustic startle response (ASR). The PPI deficit and increased ASR were resistant to antipsychotic treatment. The PPI deficit was not a consequence of impaired hearing measured by auditory brain stem responses. The Df(h22q11)/+ mice also displayed increased amplitude of loudness-dependent auditory evoked potentials. Prefrontal cortex and dorsal striatal elevations of the dopamine metabolite DOPAC and increased dorsal striatal expression of the AMPA receptor subunit GluR1 was found. The Df(h22q11)/+ mice did not deviate from wild-type mice in a wide range of other behavioural and biochemical assays. **Limitations:** The 22q11.2 microdeletion has incomplete penetrance in humans, and the severity of disease depends on the complete genetic makeup in concert with environmental factors. In order to obtain more marked phenotypes reflecting the severe conditions related to 22q11.2DS it is suggested to expose the Df(h22q11)/+ mice to environmental stressors that may unmask latent psychopathology. **Conclusion:** The Df(h22q11)/+ model will be a valuable tool for increasing our understanding of the etiology of schizophrenia and other psychiatric disorders associated with the 22q11.2DS.

## Introduction

The 22q11.2 hemizygous microdeletion confers very high risk for neurodevelopmental disorders, including autism and schizophrenia (22q11.2 deletion syndrome [22q11.2DS]). The estimated prevalence is approximately 1 in 2000.<sup>1</sup> The International Consortium on Brain and Behaviour in 22q11.2 has recently reported the cumulated prevalence of schizophrenia to be 24% in adolescence and 41% in adulthood.<sup>2</sup> Studies of patients with schizophrenia find that 22q11.2 deletion accounts for approximately 0.3% of the cases.<sup>3</sup>

Despite massive efforts there is still no coherent understanding of the etiology of schizophrenia — a highly heritable heterogeneous disorder with strong environmental influence.<sup>4,5</sup>

Several neurotransmitters are implicated in the disorder: glutamate,<sup>6</sup>  $\gamma$ -aminobutyric acid (GABA),<sup>7</sup> dopamine (DA)<sup>8</sup> and acetylcholine signalling<sup>9</sup> have all been highlighted in the disease etiology and manifestation. The cognitive impairment and negative symptomatology have been related to dysfunction in regulation of glutamate–GABA transmission leading to excitatory–inhibitory imbalances.<sup>7</sup> Like in individuals with schizophrenia,<sup>10,11</sup> cognition<sup>12</sup> and information processing is disrupted in children with 22q11.2 deletion, in whom schizophrenia has not (yet) developed.<sup>13,14</sup> Given the highly increased risk for schizophrenia and the phenotypic overlap between schizophrenia and the 22q11.2DS, studying the consequence of the 22q11.2 deletion provides a unique opportunity to add to the understanding of the

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etiology of schizophrenia and other related psychiatric disorders, which eventually may lead to novel drugs targeting the core of the disease.

Five transgenic mouse models of the 22q11.2DS have been generated by different research groups (Table 1). These models have recently been reviewed by Hiroi and colleagues,<sup>20</sup> who also provide an overview of key genes in the 22q11 region. Inconsistent results have been reported from assays addressing cognitive functions relevant to schizophrenia (unpublished observations), whereas studies of other symptom domains have been more consistent (Table 1). Decreased

prepulse inhibition (PPI) and increased acoustic startle response (ASR) have been consistently observed (LgDel,<sup>16</sup> Df1/+<sup>17</sup> and Df(16)A+/-<sup>18</sup>). However, hearing was recently shown to be impaired in the Df1/+ mouse,<sup>19</sup> raising the possibility that hearing loss rather than altered sensorimotor gating might underlie PPI deficits in 22q11DS mouse models.

The LgDel,<sup>16</sup> Df1/+<sup>17</sup> and Df(16)A+/-<sup>18</sup> mice used in the referred studies were noncongenic. Genetic background<sup>21</sup> and environmental factors<sup>22</sup> may influence phenotypic expression. To be able to control for these factors and ensure sufficient access to mice we have generated a new congenic mouse model

**Table 1: Summary of results for Df(h22q11)/+ and other 22q11.2DS mouse models**

Model	Df(h22q11)/+	Df(16)A+/-	LgDel	Df1/+	
Deletion	Dgcr2-Hira	Dgcr2-Hira	Dgcr2-Hira	Dgcr14-Ufd11	Znf74-Ctp
Strain	C57/Bl6N	C57/Bl6J	C57/Bl6N	Mixed C57/Bl6C-/C-; 129S5/SvEvBrd	129SvEvTac or mixed 129SvEvTac
Phenotype; measure					
PPI					
Pre-puberty	↓	—	—	—	—
Post-puberty	↓	↓ <sup>18</sup>	↓ <sup>16</sup>	↓ <sup>17</sup>	↑ <sup>15</sup>
Clozapine or haloperidol challenges	x	—	—	—	—
Acoustic startle					
Pre-puberty	↑	—	—	—	—
Post-puberty		x <sup>(17)</sup>	↑ <sup>16</sup>	↑ <sup>17</sup>	x <sup>(15)</sup>
Clozapine or haloperidol challenges	x	—	—	—	—
Stimulant LMA					
PCP					
Pre-puberty	x	—	—	—	—
Post-puberty	↑	—	—	—	—
Ketamine (post-puberty)	↑	—	—	—	—
Electrophysiology					
Prefrontal spike frequency					
Baseline	x	—	—	—	—
Gabazine challenge	x	—	—	—	—
Cortical low frequency oscillation					
Baseline	x	—	—	—	—
PCP challenge	x	—	—	—	—
LDAEP	↑	—	—	—	—
ABR	x	—	—	↓ <sup>19</sup>	—
Biochemistry					
hPLC (PFC and DStr)					
DOPAC	↑	—	—	—	—
DA, 5-HT, NA, 5-HIAA, HVA	x	—	—	—	—
DStr microdialysis					
Baseline DA, DOPAC, Glu, HVA	x	—	—	—	—
Veratridine, nomifensine, PCP challenge	x	—	—	—	—
Western analysis (PFC, DStr, Hipp)					
Synaptic (PSD-95, SYP, Syn1, Drebrin, geophysin)	x	—	—	—	—
Cell type (NeuN, GFAP)	x	—	—	—	—
GABA (GABAA $\alpha$ 1, KCC2, VGAT, GAD65/67)	x	—	—	—	—
Glutamate (GluR2, NR2A, NR2B, VGluT1)	x	—	—	—	—
DStr GluR1		—	—	—	—

5-HIAA = 5-hydroxyindoleacetic acid; 5-HT = 5-hydroxytryptamine; ABR = auditory induced brainstem response; DA = dopamine; DOPAC = dihydroxyphenylacetic acid; DStr = dorsal striatum; GABA =  $\gamma$ -aminobutyric acid; Hipp = hippocampus; HVA = homovanillic acid; LDAEP = loudness dependence auditory evoked potentials; LMA = locomotor activity; NA = noradrenaline; PCP = phencyclidine; PFC = prefrontal cortex; PPI = prepulse inhibition; ↓ = decreased; ↑ = increased; x = no effect; () = did not replicate.

(Df(h22q11)/+) of the 22q11.2DS as part of a large multisite collaboration. Here we report on an extensive characterization of these mice using a set of assays related to the pathophysiology of schizophrenia. The Df(h22q11)/+ mice display phenotypes relevant for modelling aspects of schizophrenia-related symptoms, including increased postpubertal *N*-methyl-D-aspartate (NMDA) antagonist sensitivity and age-independent PPI and ASR deficits. These impairments were not a result of impaired hearing. The Df(h22q11)/+ mice also display increased amplitude of loudness-dependent auditory evoked potentials (LDAEP), prefrontal cortex (PFC) and dorsal striatal elevations of the DA metabolite dihydroxyphenylacetic acid (DOPAC), and increased dorsal striatal expression of the AMPA receptor subunit GluR1. The Df(h22q11)/+ mice did not deviate from wild-type (WT) mice in a range of other behavioural and biochemical assays.

## Methods

### Animals

The Df(h22q11)/+ mouse line was generated by TaconicArtemis. Animals were bred by mating WT C57BL/6N females with hemizygotic Df(h22q11)/+ males to avoid any placental or maternal care effects of the deletion. Animals were weaned at 3 weeks, and tail biopsies were collected for polymerase chain reaction (PCR)-based genotyping. Mice were group-housed (2 WT mice and 2 hemizygotes from the same litter per cage) under controlled laboratory conditions (12-h light-dark cycle, 20 ± 2°C, 30%–70% humidity) in standard mouse cages with sawdust bedding, environmental enrichment (plastic house and paper for nesting), and food and water available ad libitum. After surgery mice were single-housed (see Appendix 1, available at [jpn.ca](http://jpn.ca)). All experiments were carried out using littermate controls. The experiments used 27 cohorts ( $n = 12$ –95) of male mice aged 6–26 weeks (Appendix 1, Fig A2). The selection of cohorts and animals for individual experiments was not randomized. All studies were carried out in accordance with the local legislation based on the European Union regulation (directive 2010/63 of Sept. 22, 2010) and UK Animals (Scientific Procedures) Act of 1986. The studies were approved by the Barcelona School of Medicine Institutional Animal Care and Use Committee or the Danish National Committee for Ethics in Animal Experimentation.

### Drugs

Clozapine (obtained from Novartis) was dissolved in 0.1 M of hydrochloride and diluted with saline. Gabazine (SR95531, obtained from Sigma-Aldrich) was dissolved in 0.2 M of NaCl. Haloperidol (obtained from Sigma) was dissolved in 0.1 M of tartaric acid and diluted with saline. Phencyclidine (PCP) hydrochloride (synthesized by Lundbeck) was dissolved in 0.1 M of methanesulfonic acid and diluted with saline. (S)-(+)-ketamine (obtained from Sigma Aldrich) was dissolved in 5% (w/v) glucose solution at a concentration of 10 mg/mL.

For the microdialysis studies, veratridine and PCP were purchased from Sigma-Aldrich and nomifensine from Tocris. Veratridine was dissolved in DMSO (5 mM) and nomifensine (1 mM) in artificial cerebrospinal fluid (aCSF).

### Basal characterization

Animals were characterized in the hot plate, rotarod, beam-walk, bright open field, locomotor activity and elevated plus maze.

### Prepulse inhibition of the ASR

The procedure has been extensively described elsewhere.<sup>23</sup> Six cohorts of animals were tested without drug treatment (age 6–21 wk). Two cohorts were treated with either clozapine (10 mL/kg administered subcutaneously at age 13 wk) or haloperidol (10 mL/kg administered subcutaneously at age 15 wk).

### NMDA receptor antagonist-induced hyperactivity

Following a 60-min habituation phase to a novel environment, Df(h22q11)/+ and WT littermates were treated with PCP or S(+)-ketamine (10 mL/kg administered subcutaneously), and their activity was monitored for a further 60 min.

### Anatomy

We conducted gross brain characterization, including assessment of brain weight, ventricular size, hippocampal structure, myelination, cortical thickness and layer composition, using cortical NeuN and parvalbumin immunoreactivity and solochrome staining.

### Western blotting

We carried out a targeted screen (Peggy-Simple western blotting) for proteins in the PFC, sorsal striatum and hippocampus. Markers showing significant changes across genotypes were replicated twice. The first replication used new dilutions from each of the 11 samples per genotype separately, whereas the second replication used 1 pooled sample for each genotype, which was run twice.

### Tissue content measured using high-performance liquid chromatography

We collected and analyzed PFC and dorsal striatal tissue content of DA, DOPAC, homovanillic acid (HVA), noradrenaline (NA), 5-hydroxytryptamine (5-HT) and 5-hydroxyindoleacetic acid (5-HIAA) as previously described.<sup>24</sup>

### Microdialysis in freely moving animals

We assessed the functional state of dorsal striatal DA transmission as previously described.<sup>25</sup> Briefly, after collection of baseline DA dialysate fractions, extracellular DA levels were measured following reverse dialysis by application of the depolarizing agent veratridine (50 mM, local 20-min pulse) and the DA/NA transporter uptake inhibitor nomifensine (50 mM, local application during 8 fractions). We subcutaneously administered PCP (2.5 + 2.5 mg/kg) systemically in the same animals the following day. Microdialysis measures were taken +0.5 mm anteroposterior (AP), -1.7 mm medial lateral (ML) and -4.5 mm dorsal ventral (DV) from Bregma.<sup>26</sup>

## Electrophysiology

### Loudness dependent auditory evoked potentials

Auditory stimuli (white noise, 5 ms duration with 1 ms rise and fall) were presented to awake freely moving mice at different intensity levels (60, 70, 80, 90 and 100 dB) with a 6-s interstimulus interval. We obtained 20-min electroencephalography (EEG) recordings during auditory stimulation. Grand average AEPs from the auditory cortex (AuC) were constructed consisting of a 100-ms prestimulus baseline and a 900-ms poststimulus interval. The peak N1 and P2 amplitudes were determined as the most negative deflection 10–25 ms poststimulus and the most positive deflection 22–65 ms poststimulus, respectively. Peak-to-peak N1/P2 amplitudes were determined at each intensity. We calculated linear regression between peak-to-peak N1/P2 amplitude and sound intensity, and the LDAEP was determined as the mean slope of the linear regression.

### Auditory-induced brainstem response

Auditory-induced brainstem responses (ABRs) were recorded with auditory stimulation to the left and right ears of anaesthetized animals with hollow ear bars guiding sound waves to the external auditory meatus. Three subdermal electrodes were inserted; the active electrode was placed at the vertex, the reference electrode was placed at the ear being tested, and the ground electrode was placed near the opposite ear. We recorded the ABRs during a series of click stimuli (duration: 50  $\mu$ s, rate: 20 Hz/s, intensity: 30–100 dB in 5-dB steps) with 500 repeats per intensity level. Stimuli were presented to 1 ear, with the nonstimulated ear being blocked with an earplug. At each intensity ABRs were averaged and analyzed within a 7-ms poststimulus window. We identified ABR thresholds by visual inspection blinded to the genotype for both left and right ears and defined the thresholds as the lowest intensity evoking a deflection of the ABR wave. The ABR protocol used was modified from that used by Fuchs and colleagues.<sup>19</sup>

### Low-frequency cortical oscillation

Medial prefrontal cortical (mPFC) low-frequency oscillation (LFO) power was assessed at baseline and following systemic PCP treatment (10 mg/kg administered subcutaneously) as previously described.<sup>27</sup> Recordings were made at +2.1 mm AP, –0.2 to –0.4 mm ML and –1 to –2.5 mm DV from Bregma.<sup>26</sup>

### Medial PFC GABAA receptor function

We assessed mPFC GABAA receptor (GABAAR) function by examining discharge rates of putative pyramidal neurons in control conditions and during local application of the GABAAR antagonist gabazine as previously described.<sup>28</sup> Recordings were made at +2.1 mm AP, –0.2 to –0.4 mm ML and –1 to –2.5 mm DV from Bregma.<sup>26</sup>

### Statistical analysis

We performed our statistical analyses using SigmaPlot version 11.2 (Systat Software Inc.) or SPSS version 21.0 (IBM Corp). Data were analyzed using either 2-way analysis of

variance (ANOVA), 2-way repeated-measures ANOVA, or Student *t* test. We conducted post hoc tests using the Holm–Sidak method, and for the *t* test we used the Mann–Whitney rank sum test if the normality (Shapiro–Wilk) or equal variance (Levene) test failed.

No adjustments were made for multiple testing beyond the post hoc tests. Data were visually inspected for outliers. No obvious outliers were identified, and all data were included in the analysis. We had only a few missing values, which we handled in SigmaPlot using a general linear model.

## Results

### Generation of mice and basic phenotyping

The Df(h22q11)/+ mouse line was generated by deletion of the human 22q11.2DS orthologous genomic region on mouse chromosome 16 (Fig. 1A and Appendix 1, Fig. A1). Expression levels of deleted genes in cortical areas were examined using microarray analysis (Fig. 1B). Except for *Tssk1*, *Vpreb2*, and *Cdc45*, expression of all detected genes in the deleted region were significantly reduced to roughly 50% of the expression in WT mice. Absolute expression levels assessed by RNAseq analysis of cortex in WT mice showed that *Tssk1*, *Vpreb2*, and *Cdc45* along with the undetected *Tssk2* and *Gsc2* genes were very lowly expressed, suggesting that the apparent unchanged expression of these genes are due to very low signal-to-noise ratio in the microarray detection (data not shown). Expression of the flanking gene *Car15* was also reduced by 50% in Df(h22q11)/+ mice, whereas expressions of other flanking genes were unaltered. Only around 40% of newborn pups were hemizygous, indicating reduced conception or intrauterine survival (Df(h22q11)/+ fraction ~0.40,  $p = 0.042$ ; Appendix 1, Fig. A3). In the studies that followed, hemizygous males were compared with WT male littermates.

Overall, Df(22q11.2)/+ mice appeared healthy with grossly normal behaviour, including pain sensitivity (hotplate: 50–56°C), motor function (beam-walking, locomotor activity, rotarod) and anxiety response (bright open field, elevated plus maze; Appendix 1, Fig. A3), and with normal brain weight and gross morphology (cortical layer composition, hippocampal structures, parvalbumin positive interneuron counts and myelin patterns; Appendix 1, Fig. A3 and A4).

### Behaviour

#### Prepulse inhibition

We found that PPI was significantly reduced in Df(h22q11)/+ mice (Fig. 2A). Data were collapsed across prepulse intensities, as there was no genotype  $\times$  prepulse intensity (5, 10, 15 dB) interaction. The PPI impairment was stable during development and observed consistently in 6 independent cohorts of 6- to 21-week-old animals. The impairment was resistant to treatment with haloperidol (Fig. 2B) and clozapine (Fig. 2C), representing typical and atypical antipsychotics, respectively. Clozapine impaired PPI in the WT mice at the highest dose tested (2 mg/kg).

**Acoustic startle response**

Df(h22q11)/+ mice had increased ASR. This was observed in 6 independent cohorts of 6- to 21-week-old animals (Fig. 2D). Haloperidol did not attenuate the increased ASR (Fig. 2E). Clozapine decreased ASR in both WT and Df(h22q11)/+ mice. However, no genotype × treatment interaction effect was observed (Fig. 2F).

**PCP-induced hyperactivity**

Df(h22q11)/+ mice showed increased activity to PCP-induced locomotor activity in an age-dependent manner. The hyperreactivity was seen in 9-week-old but not 7-week-old mice (Fig. 2G).

**S-(+)-ketamine-induced hyperactivity**

Df(h22q11)/+ mice at 26 weeks of age showed hyperreactivity to ketamine-induced locomotor activity (Fig. 2H).

*Biochemistry*

**Tissue content**

Df(h22q11)/+ mice had increased levels of DOPAC in the PFC (Fig. 3A) and dorsal striatum (Fig. 3B), whereas the levels of DA, HVA, NA, 5-HT and 5-HIAA were unaffected.

**Microdialysis in freely-moving animals**

Df(h22q11)/+ and WT mice did not differ in baseline dorsal striatal DA, DOPAC, HVA or glutamate levels (Fig. 3C). Local administration of the sodium channel opener veratridine (Fig. 3D) and the DA/NE reuptake inhibitor nomifen-sine (Fig. 3E) increased DA equally in Df(h22q11)/+ and WT mice. Similarly, PCP increased dorsal striatal DA equally in Df(h22q11)/+ and WT mice following both the first and second PCP challenge (Fig. 3F).

**Western blotting**

Df(h22q11)/+ mice showed decreased PFC NeuN and increased dorsal striatal GluR1 levels (Fig. 3G). In a replication study using pooled samples from WT and Df(h22q11)/+ mice, respectively, the dorsal striatal GluR1 increase was confirmed while the PFC NeuN signal failed to replicate. There were no effects of genotype on levels of the 14 additional markers measured (Table 2).

*Electrophysiology*

**Auditory induced brainstem response threshold**

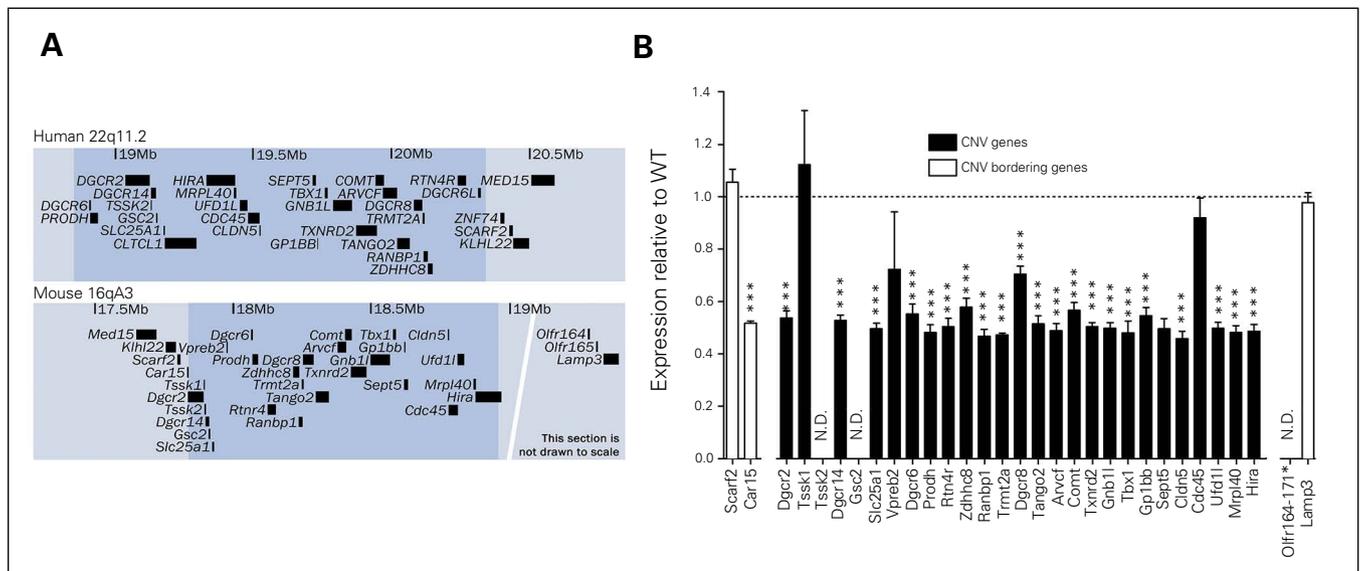
Df(h22q11)/+ and WT mice did not differ in ABR threshold (Fig. 4A). The ABR threshold did not exceed the previously defined hearing deficit criterion (55 dB SPL).<sup>19</sup>

**Loudness dependent auditory evoked potentials**

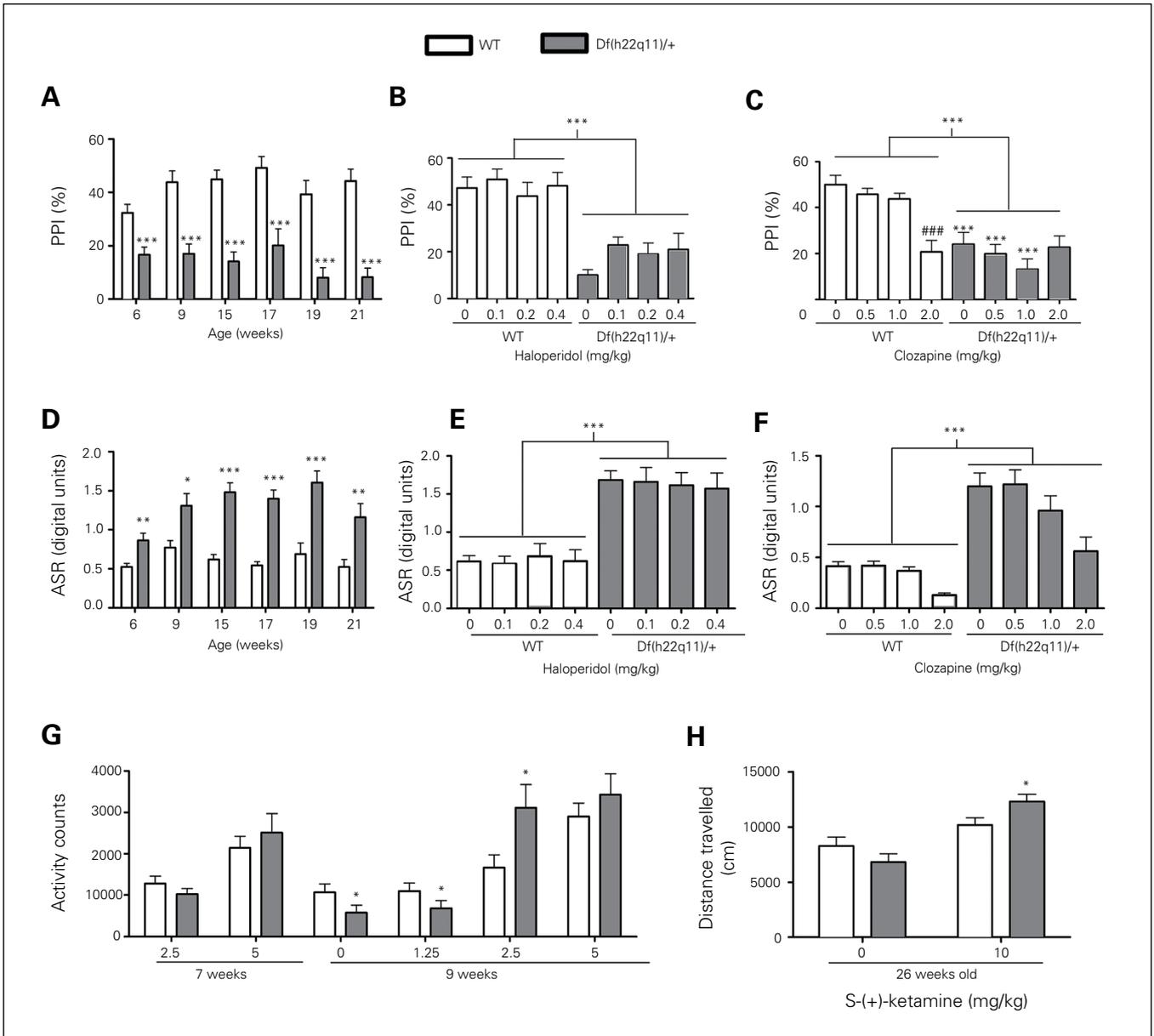
The Df(h22q11)/+ mice showed increased LDAEPs in AuC (Fig. 4D and E). There was a significant increase in P1/N1 and N1/P2 amplitudes. The Df(h22q11)/+ mice showed increased LDAEP at higher sound intensities, and the N1/P2 amplitudes were increased at 90 dB ( $p < 0.001$ ) and 100 dB ( $p < 0.001$ ). Similar results were generated for other cortical brain regions, with differences in N1 amplitude as the main contributing component (data not shown). There was no effect of genotype on latency at any sound intensity (data not shown).

**Medial PFC LFOs**

There was no difference between Df(h22q11)/+ and WT mice on baseline or PCP-induced attenuation of cortical LFOs (Fig. 4B). Administration of PCP decreased cortical LFOs.



**Fig. 1:** Construct similarities in Df(h22q11)/+ and human deletion carriers. (A) Overview of the deleted region (shaded) in humans (22q11.2) and the corresponding orthologous region in mice (16qA3). The maps are based on human library GRCh38/hg38 and mouse library GRCm38/mm10 from the University of California, Santa Cruz (UCSC) database. Only annotated RefSeq sequences are shown for clarity. (B) Median normalized microarray analysis of relative cortical expression of gene products from deleted segment in Df(h22q11)/+ mice compared with wild-type (WT) littermates. \*Significant difference between WT and Df(h22q11)/+ mice (\*\* $p < 0.001$ ). CNV = copy number variation; N.D. = not detected.



**Fig. 2:** Behavioural characterization of Df(h22q11)/+ mice and wild-type (WT) littermates. **(A)** Df(h22q11)/+ mice showed decreased prepulse inhibition (PPI). The PPI deficit was reproduced in 6 independent cohorts of mice 6–21 weeks old ( $n = 12$ –24/group). We conducted a  $t$  test for each week (week 6:  $t_{45} = 3.6$ ,  $p < 0.001$ ; week 9:  $t_{30} = 4.8$ ,  $p < 0.001$ ; week 15:  $t_{29} = 6.3$ ,  $p < 0.001$ ; week 17:  $t_{22} = 3.9$ ,  $p < 0.001$ ; week 19:  $t_{22} = 4.9$ ,  $p < 0.001$ ; week 21:  $t_{22} = 6.4$ ,  $p < 0.001$ ). **(B)** Haloperidol did not rescue the PPI deficit ( $n = 12$ /group, 2-way analysis of variance [ANOVA], genotype:  $F_{1,94} = 69.1$ ,  $p < 0.001$ ; dose:  $F_{3,94} = 1.02$ ,  $p = 0.38$ ; genotype  $\times$  dose:  $F_{3,94} = 0.59$ ,  $p = 0.63$ ). **(C)** Clozapine did not rescue the PPI deficit ( $n = 13$ /group, 2-way ANOVA, genotype:  $F_{1,87} = 46.3$ ,  $p < 0.001$ ; dose:  $F_{3,87} = 4.87$ ,  $p = 0.004$ ; genotype  $\times$  dose:  $F_{3,87} = 6.14$ ,  $p < 0.001$ ). **(D)** Df(h22q11)/+ mice had increased acoustic startle response (ASR). The increased ASR was found in 6 independent cohorts of mice 6–21 weeks old ( $n = 12$ –24/group). We conducted a  $t$  test for each week (week 6:  $T_{n23,24} = 418$ ,  $p = 0.004$ ; week 9:  $T_{n16,16} = 204$ ,  $p = 0.025$ ; week 15:  $T_{n15,16} = 340$ ,  $p < 0.001$ ; week 17:  $T_{n12,12} = 79$ ,  $p < 0.001$ ; week 19:  $t_{22} = -4.51$ ,  $p < 0.001$ ; week 21:  $T_{n12,12} = 101$ ,  $p = 0.005$ ). **(E)** Haloperidol did not attenuate the increased ASR ( $n = 12$ /group, 2-way ANOVA, genotype:  $F_{1,94} = 88.5$ ,  $p < 0.001$ ; dose:  $F_{3,94} = 0.060$ ,  $p = 0.98$ ; genotype  $\times$  dose:  $F_{3,94} = 0.13$ ,  $p = 0.95$ ). **(F)** Clozapine did not attenuate the increased ASR ( $n = 13$ /group, 2-way ANOVA, genotype:  $F_{1,87} = 83.1$ ,  $p < 0.001$ ; dose:  $F_{3,87} = 9.35$ ,  $p < 0.001$ ; genotype  $\times$  dose:  $F_{3,87} = 1.49$ ,  $p = 0.22$ ). **(G)** Age-dependent hypersensitivity to phencyclidine (PCP)-induced locomotion in Df(h22q11)/+ mice ( $n = 12$ /group). We conducted a  $t$  test for each week (week 7:  $t_{22} = 1.10$ ,  $p = 0.28$  at 2.5 mg/kg and  $t_{22} = -0.68$ ,  $p = 0.50$  at 5.0 mg/kg; week 9:  $T_{n11,12} = 90$ ,  $p = 0.011$  for vehicle;  $T_{n12,12} = 187$ ,  $p = 0.035$  at 1.25 mg/kg;  $t_{22} = -2.33$ ,  $p = 0.029$  at 2.5 mg/kg and  $t_{22} = -0.91$ ,  $p = 0.37$  at 5.0 mg/kg). **(H)** Hypersensitivity to ketamine-induced locomotion in Df(h22q11)/+ mice ( $n = 13$ –16/group, 2-way ANOVA, genotype:  $F_{1,54} = 0.23$ ,  $p = 0.64$ ; dose:  $F_{11,54} = 25.6$ ,  $p < 0.001$ ; genotype  $\times$  dose:  $F_{1,54} = 5.95$ ,  $p = 0.018$ ; post hoc test within ketamine 10 mg/kg:  $t = 2.05$ ,  $p = 0.044$ ). Data are presented as mean  $\pm$  standard errors of the mean (SEM). \*Significant differences between WT and Df(h22q11)/+ mice (\* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ ). #Significant difference within genotype relative to vehicle (# $p < 0.05$ , ### $p < 0.001$ ).

**Medial PFC GABAAR function**

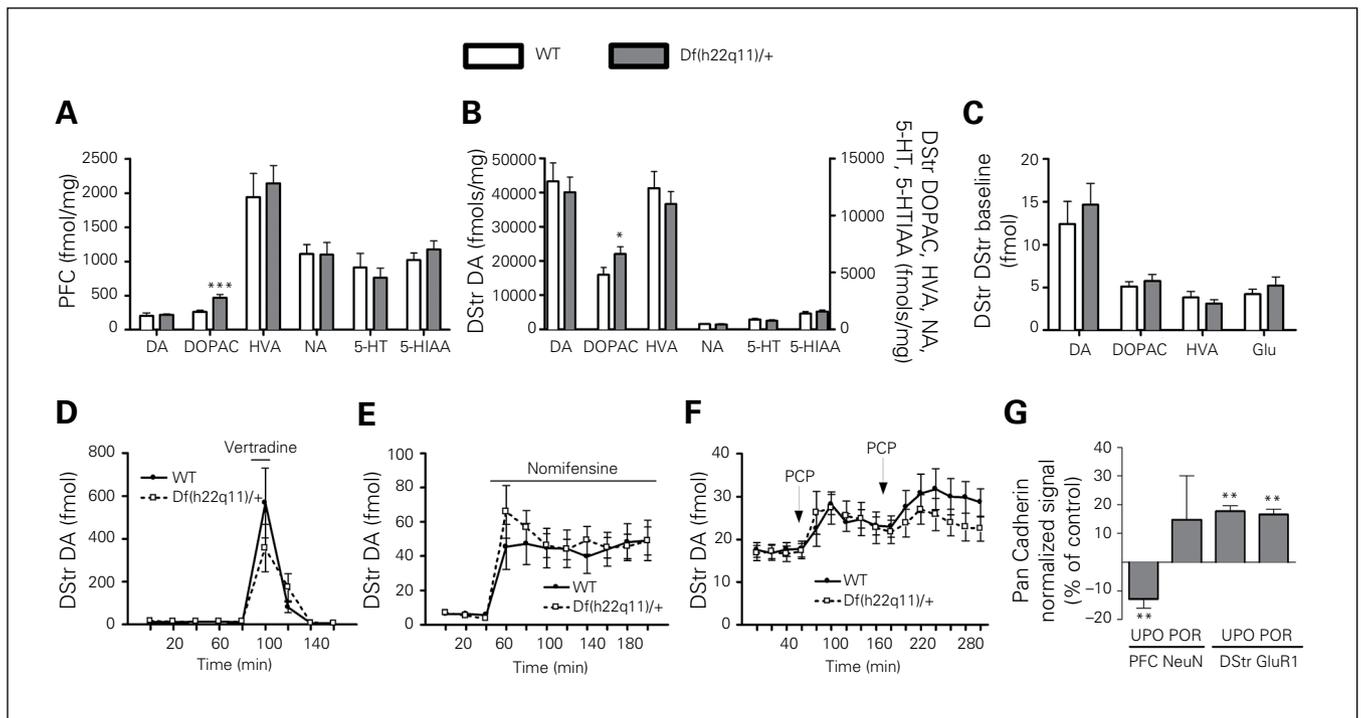
There was no difference between Df(h22q11)/+ and WT mice on baseline or GABAAR antagonist-induced elevation of pyramidal neuron spike frequency (Fig. 4C).

**Discussion**

In this paper we report on the generation and characterization of a new 22q11DS mouse model with a focus on assays relevant for the pathophysiology of schizophrenia. The Df(h22q11)/+ mice had age-independent and antipsychotic-resistant gating deficits measured by PPI. They had increased ASR and increased amplitude of LDAEP. Also, the Df(h22q11)/+ mice exhibited increased NMDAR antagonist-induced locomotion, but in contrast to PPI, this effect was observed only after puberty. The 22q11.2 deletion resulted in increased dorsal striatal and PFC DOPAC levels, as would be

expected by the deletion of the *Comt* gene. Among other markers explored (Fig. 3 and Table 2), we saw a reduction of dorsal striatal GluR1 expression.

Prepulse inhibition deficits and ASR changes have previously been reported in other 22q11DS mouse models.<sup>16-18</sup> We confirmed these findings in our model and further examined the age dependency, response to antipsychotic treatment and possible hearing confounds. We found that the PPI and ASR changes in Df(h22q11)/+ mice are robust (observed in 6 independent cohorts), are present before puberty and persist across ages. This effect mirrors the reduced PPI found in human 22q11.2 deletion carriers, which precedes onset of puberty and the potential development of schizophrenia symptoms.<sup>13,14</sup> Chronic middle ear infection and elevated click-response ABR threshold have been observed in the Df1/+ mouse model<sup>19</sup> raising the possibility that hearing loss at low intensities rather than sensorimotor gating deficits



**Fig. 3:** Biochemical measures in Df(h22q11)/+ mice and wild-type (WT) littermates. **(A)** Whole tissue content in the prefrontal cortex (PFC). The level of 3,4-dihydroxyphenylacetic acid (DOPAC) was increased in Df(h22q11)/+ mice ( $n = 8-10$ /group). We conducted a  $t$  test for each analyte (dopamine [DA]:  $t_{16} = -0.22, p = 0.83$ ; DOPAC:  $t_{16} = -3.85, p = 0.001$ ; homovanillic acid [HVA]:  $t_{16} = -0.48, p = 0.64$ ; noradrenaline [NA]:  $T_{n8,10} = 81, p = 0.69$ ; 5-hydroxytryptamine [5-HT]:  $T_{n8,10} = 83, p = 0.56$ ; 5-hydroxyindoleacetic acid [5-HIAA]:  $t_{16} = -0.93, p = 0.38$ ). **(B)** Dorsal striatum (DStr) whole tissue content. The level of DOPAC was increased in Df(h22q11)/+ mice ( $n = 9-15$ /group). We conducted a  $t$  test for each analyte (DA:  $t_{27} = 0.45, p = 0.66$ ; DOPAC:  $T_{n14,15} = 156, p = 0.020$ ; HVA:  $T_{n14,15} = 215, p = 0.84$ ; NA:  $T_{n15,15} = 238, p = 0.85$ ; 5-HT:  $t_{28} = 0.91, p = 0.37$ ; 5-HIAA:  $T_{n15,15} = 198, p = 0.16$ ). **(C)** No genotype effect was detected in the microdialysis of the DStr at baseline ( $n = 14-15$ /group). We conducted a  $t$  test for each analyte (DA:  $T_{13,15} = 203, p = 0.52$ ; DOPAC:  $t_{27} = -0.69, p = 0.50$ ; HVA:  $t_{19} = 0.98, p = 0.34$ ; glutamate [Glu]:  $T_{14,14} = 191, p = 0.58$ ). **(D)** Veratridine (VTD) increased DStr DA similarly in WT and Df(h22q11)/+ mice ( $n = 14$ /group, 2-way repeated-measures ANOVA, genotype:  $F_{1,26} = 0.24, p = 0.63$ ; time:  $F_{8,208} = 18.8, p < 0.001$ ; genotype  $\times$  time:  $F_{8,208} = 1.35, p = 0.22$ ). **(E)** Nomifensine increased DStr DA similarly in WT and Df(h22q11)/+ mice ( $n = 14$ /group, 2-way repeated-measures ANOVA, genotype:  $F_{1,26} = 0.12, p = 0.74$ ; time:  $F_{10,260} = 25.5, p < 0.001$ ; genotype  $\times$  time:  $F_{10,260} = 0.79, p = 0.64$ ). **(F)** PCP increased DStr DA similarly in WT and Df(h22q11)/+ mice ( $n = 14$ /group) following both the first (genotype:  $F_{1,28} = 0.003, p = 0.958$ ; genotype  $\times$  time:  $F_{4,112} = 0.742, p = 0.565$ ) and second PCP challenge (genotype:  $F_{1,28} = 1.632, p = 0.212$ ; genotype  $\times$  time:  $F_{5,140} = 0.432, p = 0.826$ ). **(G)** The initial Western blotting showed decreased PFC NeuN ( $t_{20} = 2.92, p = 0.009$ ) and increased DStr GluR1 expression ( $t_{20} = 2.99, p = 0.007$ ) in Df(h22q11)/+ mice ( $n = 11$ /group). However, the NeuN decrease failed to replicate when samples were pooled. Data are presented as means + standard errors of the mean (SEM). \*Significant differences between WT and Df(h22q11)/+ mice (\* $p < 0.05$ , \*\* $p < 0.01$ ). POR = pooled replication; UPO = unpooled.

per se underlie the PPI deficits found in the 22q11DS mouse models. We addressed this by measuring ABR thresholds and found those to be unchanged in the Df(h22q11)/+ mice. This, together with the increased amplitude of AEPs found in the LDAEP assay, indicates that hearing is not reduced in the Df(h22q11)/+ mouse. Thus, the decrease in sensorimotor gating observed in the Df(h22q11)/+ mouse is not caused by hearing loss. The contrasting ABR findings between the Df(h22q11)/+ and Df1/+ mouse models<sup>19</sup> may be due to differences in strain or environmental conditions affecting the risk of ear infection.

The PPI and ASR impairments were resistant to antipsychotic treatment (haloperidol and clozapine), which is in overall agreement with the effect of these drugs in the clinical setting. In patients with schizophrenia, classical antipsychotics working through D2/D3 receptor blockade reduce the psychotic symptoms without affecting PPI deficits.<sup>29,30</sup> The findings for the new generation of antipsychotics with multi-receptor profiles are more mixed,<sup>29,31,32</sup> and a cross-sectional study by Kumari and colleagues<sup>33</sup> provided indirect evidence of clozapine being superior to haloperidol for PPI deficits in patients with schizophrenia, which is not reflected by the present data from the Df(h22q11)/+ mice.

Similar to the Df1/+ mouse,<sup>34</sup> we found that Df(h22q11)/+ mice displayed increased locomotor activity in response to the NMDAR antagonists PCP and ketamine. Interestingly, the effect was age-dependent and only was observed after puberty (9- but not 7-week-old animals). As schizophrenia is characterized by onset of overt symptoms during or after puberty<sup>35</sup> and glutamatergic dysfunctions,<sup>36,37</sup> our data suggest that the Df(h22q11)/+ mouse may model aberrant NMDAR-related neurodevelopment trajectories associated with the disorder. The prepubertal and age-independent PPI deficits found in the Df(h22q11)/+ mice suggest that these sensorimotor gating deficits are due

to biological perturbation other than what drives the NMDAR antagonist phenotype.

Biochemical assays revealed that the Df(h22q11)/+ mice had elevated PFC and dorsal striatal tissue content of the DA metabolite DOPAC. This phenotype may be explained by the haploinsufficiency for COMT, which catalyzes the demethylation of DA. The role of COMT is well established in low-DAT density brain regions, such as the PFC,<sup>38,39</sup> whereas there are contrasting findings in the striatum.<sup>40,41</sup> The present data also support a role of COMT in striatal DA function. Despite the reduced expression of COMT and consequent increase in DOPAC we saw no changes in tissue content of DA and HVA, NA or 5-HT. The Df(h22q11)/+ mice also showed normal dorsal striatal DA release following PCP administration and local application of veratridine or nomifensine. Dopamine release is involved in, but temporally dissociated from, NMDA antagonist-induced locomotor activity.<sup>42</sup> The observed hypersensitivity to NMDAR blockade is likely not caused by hypersensitivity of the DA system in the Df(h22q11)/+ mice, as release of DA is unchanged. This is further supported by the lack of locomotor hyperresponsivity to amphetamine in these mice (unpublished data). GluR1 was the only glutamatergic-related marker (GluR2, NR2A, NR2B, or VGluT1) explored that showed differential expression, and we did not observe any changes in GABA-related markers (GABAA  $\alpha$ 1, KCC2, VGAT, or GAD 65/67). Furthermore, PFC GABAergic–glutamatergic interaction measured as GABAAR antagonism-induced pyramidal cell discharge rates was not affected, and measures of PCP-induced attenuation of LFOs showed preserved function of cortical neuronal networks in the Df(h22q11)/+ mice. Thus, expression of most receptors for the major transmitters is not altered, and the mechanism responsible for increased behavioural responsiveness to NMDAR blockade is unknown, which should be investigated further in future studies.

**Table 2: Protein level in Df(h22q11)/+ mice and wild-type littermates**

Type	Marker	Function	PFC	DStr	Hipp
Synaptic	PSD-95	Scaffolding protein at excitatory post synaptic densities	x	x	x
	Synaptophysin	Synaptic vesicle protein	x	x	x
	Synapsin 1	Phosphoprotein associated with surface of synaptic vesicles	x	x	x
	Drebrin	Actin-binding protein involved in neuronal /spine growth	x	x	x
	Gephyrin	Scaffolding protein in inhibitory synaptic densities	x	x	x
Cell type	NeuN	Nuclear antigen. Biomarker of neurons	(↓)	x	x
	GFAP	Intermediate filament in astrocytes	x	x	x
Glutamate	GluR1	AMPA receptor subunit	x	↑	x
	GluR2	AMPA receptor subunit	x	x	x
	NR2A	NMDA receptor subunit	x	x	x
	NR2B	NMDA receptor subunit	x	x	x
	VGluT1	Vesicular glutamate transporter at vesicle membrane	x	x	x
GABA	GABAA $\alpha$ 1	GABA <sub>A</sub> receptor subunit	x	x	x
	KCC2	Neuron specific potassium-chloride transporter	x	x	x
	VGAT	Vesicular GABA transporter found on synaptic vesicle membrane	x	x	x
	GAD 65/6	Enzymes catalyzing decarboxylation of glutamate to GABA	x	x	x

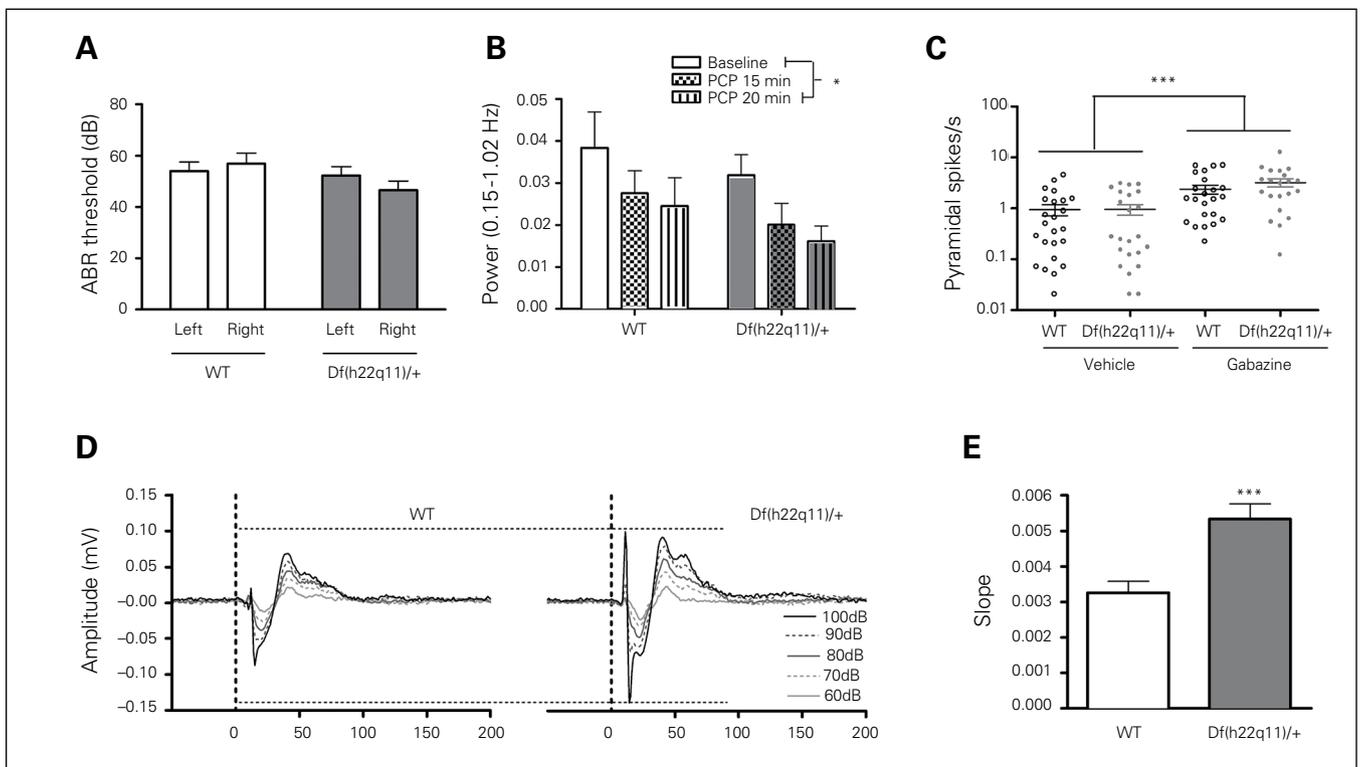
DStr = dorsal striatum; GABA =  $\gamma$ -aminobutyric acid; Hipp = hippocampus; NMDA = *N*-methyl-D-aspartate; PFC = prefrontal cortex; ↓ = decreased; ↑ = increased; x = no effect; () = did not replicate.

In line with the increased ASR, the amplitude of AEPs were increased in *Df(h22q11)/+* mice. The amplitude of AEPs has been found to be decreased in patients with schizophrenia,<sup>43</sup> whereas mixed results have been found for LDAEP. Some studies have reported that LDAEP was reduced in patients with schizophrenia, but these studies were not designed to account for difference in symptomatology.<sup>44,45</sup> In a recent study, Wyss and colleagues<sup>46</sup> found increased LDAEP in patients with schizophrenia with predominant negative symptoms, which interestingly is also prominent in the symptomatology of patients with the 22q11.2DS<sup>47-49</sup> together with increased auditory event-related potentials.<sup>50</sup> Negative symptoms are also prominent in patients with autism, another disorder associated with the 22q11.2DS, and increased amplitude of AEPs have been reported in patients with autism-related syndromes, such as fragile X,<sup>51</sup> and in animal models of autism.<sup>52,53</sup> We have not assessed behavioural functions related to negative symptomatology in the present study. However, reports of a reduced progressive ratio response in *Df(16)A+/-* mice<sup>54</sup> supports negative symptom-like behaviour in these mice.<sup>55</sup> Hence the

*Df(h22q11)/+* mouse may also prove to be a useful model for aspects of the negative symptomatology in patients with psychiatric disorders.

*Limitations*

Genetic mouse models provide insight into the consequence of the mutation in a specific genetic and environmental context. More marked changes might have been anticipated from introducing a high-risk genetic variant comprising more than 25 genes into the mouse. However, it is important to keep in mind that the 22q11.2 microdeletion has incomplete penetrance in humans and that expression and severity of disease depends on the complete genetic makeup<sup>21</sup> in concert with environmental influences. Thus, the *Df(h22q11)/+* mouse is more appropriately thought of as a liability model rather than a disease model. This is in line with our previous findings that copy number variants conferring high risk for mental disorders significantly affect structure and function of the brain in healthy human carriers.<sup>56</sup> In order to obtain more marked phenotypes reflecting the severe conditions related



**Fig. 4:** Electrophysiological measures in *Df(h22q11)/+* and wild-type (WT) littermates. **(A)** Hearing did not differ between the *Df(h22q11)/+* mice and WT littermates ( $n = 13/\text{group}$ , 2-way repeated-measures ANOVA, genotype:  $F_{1,24} = 1.649$ ,  $p = 0.211$ ; ear:  $F_{1,24} = 0.331$ ,  $p = 0.571$ ; ear  $\times$  genotype:  $F_{1,24} = 3.570$ ,  $p = 0.071$ ). **(B)** Phencyclidine (PCP) caused similar decrease of cortical low-frequency oscillations (LFOs) in WT and *Df(h22q11)/+* mice ( $n = 9/\text{group}$ , 2-way ANOVA, genotype:  $F_{1,48} = 2.40$ ,  $p = 0.13$ ; condition:  $F_{2,48} = 3.44$ ,  $p = 0.040$ ; genotype  $\times$  condition:  $F_{2,48} = 0.013$ ,  $p = 0.99$ ). **(C)** Gabazine caused a similar increase in pyramidal spike frequencies in WT and *Df(h22q11)/+* mice ( $n = 14/\text{group}$ , 2-way ANOVA; genotype:  $F_{1,87} = 1.21$ ,  $p = 0.28$ ; treatment:  $F_{1,87} = 20.9$ ,  $p < 0.001$ ; genotype  $\times$  treatment:  $F_{1,87} = 1.12$ ,  $p = 0.29$ ). **(D)** Grand average loudness-dependent auditory evoked potentials (LDAEP) waveforms from the auditory cortex (AuC) in WT and *Df(h22q11)/+* mice ( $n = 22\text{--}23/\text{group}$ , 2-way repeated-measure ANOVA, P1/N1 genotype:  $F_{1,43} = 22.6$ ,  $p < 0.001$ ; intensity:  $F_{4,43} = 75.5$ ,  $p < 0.001$ ; genotype  $\times$  intensity:  $F_{4,43} = 13.7$ ,  $p < 0.001$ ; N1/P2 genotype:  $F_{1,43} = 9.89$ ,  $p = 0.003$ ; intensity:  $F_{4,43} = 156.4$ ,  $p < 0.001$ ; genotype  $\times$  intensity:  $F_{4,43} = 9.76$ ,  $p < 0.001$ ). **(E)** Collapsed LDAEP slopes for *Df(h22q11)/+* and WT mice ( $n = 22\text{--}23/\text{group}$ ,  $t$  test;  $t_{43} = -3.896$ ,  $p < 0.001$ ). Data are presented as means  $\pm$  standard errors of the mean. \*Significant differences ( $*p < 0.05$ ,  $***p < 0.001$ ). ABR = auditory-induced brainstem response.

to 22q11.2DS it has been suggested to combine the Df(h22q11)/+ mouse with environmental stressors, as used by Giovanoli and colleagues,<sup>57</sup> which may unmask latent psychopathology. Giovanoli and colleagues showed how unpredictable stress during puberty resulted in PPI deficits and increased sensitivity to amphetamine and MK-801 in mice exposed to prenatal immune activation. In addition, Harper and colleagues<sup>22</sup> showed how reduced social interaction in Sept5 knockout mice, one of the 22q11.2 genes, could be rescued by reducing the level of stress, clearly demonstrating a genotype × environment interaction.

Sex differences in the behaviour of children with 22q11.2DS have been reported.<sup>58</sup> Thus, it is highly relevant to explore sex differences in the mouse. In the present study only male mice were included. Pilot studies conducted in our laboratory comparing male and female Df(h22q11)/+ mice did not reveal differences in motor function, seizure threshold, anxiety, or amphetamine-induced locomotion. The studies have to be repeated and extended before firm conclusions can be made.

We observed only a few changes in protein expression. Additional synaptic fractionation might increase the resolution of the analysis, particularly with respect to synaptic proteins. However, the method used allowed us to investigate both synaptic (e.g., PSD95) and astrocytic markers (e.g., GFAP).

## Conclusion

Here we show how the Df(h22q11)/+ mouse model reflects different developmental aspects of the 22q11.2DS and schizophrenia, including changes in information processing and NMDAR antagonist responsiveness. Furthermore, the present findings extends previous reports on other 22q11.2DS mouse models by showing age-independent and antipsychotic-resistant gating deficits that are not confounded by reduced hearing. This model will be a valuable tool for further understanding the etiology of schizophrenia and other psychiatric disorders with the ultimate goal of developing novel and improved drugs.

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## References

- Shprintzen RJ. Velo-cardio-facial syndrome: 30 years of study. *Dev Disabil Res Rev* 2008;14:3-10.
- Schneider M, Debbane M, Bassett AS, et al. Psychiatric disorders from childhood to adulthood in 22q11.2 deletion syndrome: results from the International Consortium on Brain and Behavior in 22q11.2 Deletion Syndrome. *Am J Psychiatry* 2014;171:627-39.
- Rees E, Walters JT, Chambert KD, et al. CNV analysis in a large schizophrenia sample implicates deletions at 16p12.1 and SLC1A1 and duplications at 1p36.33 and CGNL1. *Hum Mol Genet* 2014; 23:1669-76.
- Harrison PJ, Weinberger DR. Schizophrenia genes within cortical neural circuits. *Mol Psychiatry* 2005;10:5.
- Ripke S, Neale BM, Corvin A, et al. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 2014;511:421-7.
- Javitt DC, Zukin SR. Recent advances in the phencyclidine model of schizophrenia. *Am J Psychiatry* 1991;148:1301-8.
- Pocklington AJ, Rees E, Walters JT, et al. Novel findings from CNVs implicate inhibitory and excitatory signaling complexes in schizophrenia. *Neuron* 2015;86:1203-14.
- Laruelle M, Abi-Dargham A, van Dyck CH, et al. Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug-free schizophrenic subjects. *Proc Natl Acad Sci U S A* 1996;93:9235-40.
- Raedler TJ, Bymaster FP, Tandon R, et al. Towards a muscarinic hypothesis of schizophrenia. *Mol Psychiatry* 2007;12:232-46.
- Elvevåg B, Goldberg TE. Cognitive impairment in schizophrenia is the core of the disorder. *Crit Rev Neurobiol* 2000;14:1-21.
- Braff D, Stone C, Callaway E, et al. Prestimulus effects on human startle reflex in normals and schizophrenics. *Psychophysiology* 1978; 15:339-43.
- Hooper SR, Curtiss K, Schoch K, et al. A longitudinal examination of the psychoeducational, neurocognitive, and psychiatric functioning in children with 22q11.2 deletion syndrome. *Res Dev Disabil* 2013;34:1758-69.
- Sobin C, Kiley-Brabeck K, Karayiorgou M. Associations between prepulse inhibition and executive visual attention in children with the 22q11 deletion syndrome. *Mol Psychiatry* 2005;10:553-62.
- Sobin C, Kiley-Brabeck K, Karayiorgou M. Lower prepulse inhibition in children with the 22q11 deletion syndrome. *Am J Psychiatry* 2005; 162:1090-9.
- Kimber WL, Hsieh P, Hirotsune S, et al. Deletion of 150 kb in the minimal DiGeorge/velocardiofacial syndrome critical region in mouse. *Hum Mol Genet* 1999;8:2229-37.
- Long JM, LaPorte P, Merscher S, et al. Behavior of mice with mutations in the conserved region deleted in velocardiofacial/DiGeorge syndrome. *Neurogenetics* 2006;7:247-57.

17. Paylor R, McIlwain KL, McAninch R, et al. Mice deleted for the DiGeorge/velocardiofacial syndrome region show abnormal sensorimotor gating and learning and memory impairments. *Hum Mol Genet* 2001;10:2645-50.
18. Stark KL, Xu B, Bagchi A, et al. Altered brain microRNA biogenesis contributes to phenotypic deficits in a 22q11-deletion mouse model. *Nat Genet* 2008;40:751-60.
19. Fuchs JC, Zinnamon FA, Taylor RR, et al. Hearing loss in a mouse model of 22q11.2 deletion syndrome. *PLoS One* 2013;8:e80104.
20. Hiroi N, Takahashi T, Hishimoto A, et al. Copy number variation at 22q11.2: from rare variants to common mechanisms of developmental neuropsychiatric disorders. *Mol Psychiatry* 2013;18:1153-65.
21. Suzuki G, Harper KM, Hiramoto T, et al. Sept5 deficiency exerts pleiotropic influence on affective behaviors and cognitive functions in mice. *Hum Mol Genet* 2009;18:1652-60.
22. Harper KM, Hiramoto T, Tanigaki K, et al. Alterations of social interaction through genetic and environmental manipulation of the 22q11.2 gene Sept5 in the mouse brain. *Hum Mol Genet* 2012;21:3489-99.
23. Fejgin K, Nielsen J, Birkenow MR, et al. A mouse model that recapitulates cardinal features of the 15q13.3 microdeletion syndrome including schizophrenia- and epilepsy-related alterations. *Biol Psychiatry* 2014;76:128-37.
24. Adell A, Sarna GS, Hutson PH, et al. An in vivo dialysis and behavioural study of the release of 5-HT by p-chloroamphetamine in reserpine-treated rats. *Br J Pharmacol* 1989;97:206-12.
25. Masana M, Bortolozzi Aa, Artigas F. Selective enhancement of mesocortical dopaminergic transmission by noradrenergic drugs: therapeutic opportunities in schizophrenia. *Int J Neuropsychopharmacol* 2011;14:53-68.
26. Paxinos G, Franklin KBJ. *The mouse brain in stereotaxic coordinates*. London: Elsevier Academic Press; 2008. p. 242.
27. Kargieman L, Santana N, Mengod G, et al. Antipsychotic drugs reverse the disruption in prefrontal cortex function produced by NMDA receptor blockade with phencyclidine. *Proc Natl Acad Sci U S A* 2007;104:14843-8.
28. Lladó-Pelfort L, Santana N, Ghisi V, et al. 5-HT1A receptor agonists enhance pyramidal cell firing in prefrontal cortex through a preferential action on GABA interneurons. *Cereb Cortex* 2012;22:1487-97.
29. Mackeprang T, Kristiansen KT, Glenthøj BY. Effects of antipsychotics on prepulse inhibition of the startle response in drug-naïve schizophrenic patients. *Biol Psychiatry* 2002;52:863-73.
30. Düring S, Glenthøj BY, Andersen GS, et al. Effects of dopamine D2/D3 blockade on human sensory and sensorimotor gating in initially antipsychotic-naïve, first-episode schizophrenia patients. *Neuropsychopharmacology* 2014;39:3000-8.
31. Duncan E, Szilagyí S, Schwartz M, et al. Prepulse inhibition of acoustic startle in subjects with schizophrenia treated with olanzapine or haloperidol. *Psychiatry Res* 2003;120:1-12.
32. Aggernaes B, Glenthøj BY, Ebdrup BH, et al. Sensorimotor gating and habituation in antipsychotic-naïve, first-episode schizophrenia patients before and after 6 months' treatment with quetiapine. *Int J Neuropsychopharmacol* 2010;13:1383-95.
33. Kumari V, Soni W, Sharma T. Normalization of information processing deficits in schizophrenia with clozapine. *Am J Psychiatry* 1999;156:1046-51.
34. Kimoto S, Muraki K, Toritsuka M, et al. Selective overexpression of Comt in prefrontal cortex rescues schizophrenia-like phenotypes in a mouse model of 22q11 deletion syndrome. *Transl Psychiatry* 2012;2:e146-16.
35. Castle D, Sham P, Murray R. Differences in distribution of ages of onset in males and females with schizophrenia. *Schizophr Res* 1998;33:179-83.
36. Olney JW, Farber NB. Glutamate receptor dysfunction and schizophrenia. *Arch Gen Psychiatry* 1995;52:998-1007.
37. du Bois TM, Huang XF. Early brain development disruption from NMDA receptor hypofunction: relevance to schizophrenia. *Brain Res Rev* 2007;53:260-70.
38. Karoum F, Chrapusta SJ, Egan MF. 3-Methoxytyramine is the major metabolite of released dopamine in the rat frontal cortex: reassessment of the effects of antipsychotics on the dynamics of dopamine release and metabolism in the frontal cortex, nucleus accumbens, and striatum by a simple two pool model. *J Neurochem* 1994;63:972-9.
39. Gogos JA, Morgan M, Luine VV, et al. Catechol-O-methyltransferase-deficient mice exhibit sexually dimorphic changes in catecholamine levels and behavior. *Proc Natl Acad Sci U S A* 1998;95:9991-6.
40. Simpson EH, Morud J, Winiger V, et al. Genetic variation in COMT activity impacts learning and dopamine release capacity in the striatum. *Learn Mem* 2014;21:205-14.
41. Yavich L, Forsberg MM, Karayiorgou M, et al. Site-specific role of catechol-O-methyltransferase in dopamine overflow within prefrontal cortex and dorsal striatum. *J Neurosci* 2007;27:10196-209.
42. Adams B, Moghaddam B. Corticolimbic dopamine neurotransmission is temporally dissociated from the cognitive and locomotor effects of phencyclidine. *J Neurosci* 1998;18:5545-54.
43. Light GA, Swerdlow NR, Rissling AJ, et al. Characterization of neurophysiologic and neurocognitive biomarkers for use in genomic and clinical outcome studies of schizophrenia. *PLoS One* 2012;7:e39434.
44. Gudlowski Y, Ozgurda S, Witthaus H, et al. Serotonergic dysfunction in the prodromal, first-episode and chronic course of schizophrenia as assessed by the loudness dependence of auditory evoked activity. *Schizophr Res* 2009;109:141-7.
45. Juckel G, Gudlowski Y, Wutzler A, et al. Loudness dependence of the auditory evoked N1/P2 component as an indicator of serotonergic dysfunction in patients with schizophrenia — a replication study. *Psychiatry Res* 2008;158:79-82.
46. Wyss C, Hitz K, Hengartner MP, et al. The loudness dependence of auditory evoked potentials (LDAEP) as an indicator of serotonergic dysfunction in patients with predominant schizophrenic negative symptoms. *PLoS One* 2013;8:e68650.
47. Schneider M, Van der Linden M, Glaser B, et al. Preliminary structure and predictive value of attenuated negative symptoms in 22q11.2 deletion syndrome. *Psychiatry Res* 2012;196:277-84.
48. Schneider M, Van der Linden M, Menghetti S, et al. Negative and paranoid symptoms are associated with negative performance beliefs and social cognition in 22q11.2 deletion syndrome. *Early Interv Psychiatry* 2015;10:1111.
49. Stoddard J, Niendam T, Hendren R, et al. Attenuated positive symptoms of psychosis in adolescents with chromosome 22q11.2 deletion syndrome. *Schizophr Res* 2010;118:118-21.
50. Rihs TA, Tomescu MI, Britz J, et al. Altered auditory processing in frontal and left temporal cortex in 22q11.2 deletion syndrome: a group at high genetic risk for schizophrenia. *Psychiatry Res* 2013;212:141-9.
51. Van der Molen MJ, Van der Molen MW, Ridderinkhof KR, et al. Auditory change detection in fragile X syndrome males: a brain potential study. *Clin Neurophysiol* 2012;123:1309-18.
52. Rotschafer S, Razak K. Altered auditory processing in a mouse model of fragile X syndrome. *Brain Res* 2013;1506:12-24.
53. Liao W, Gandal MJ, Ehrlichman RS, et al. MeCP2+/- mouse model of RTT reproduces auditory phenotypes associated with Rett syndrome and replicate select EEG endophenotypes of autism spectrum disorder. *Neurobiol Dis* 2012;46:88-92.
54. Graf R, Hanks A, Lotarski S, et al. Behavioral assessment of a mouse model of 22q11 deletion syndrome and its relevance to schizophrenia. *Biol Psychiatry* 2014;75:178S.
55. Wolf DH, Satterthwaite TD, Kantrowitz JJ, et al. Amotivation in schizophrenia: integrated assessment with behavioral, clinical, and imaging measures. *Schizophr Bull* 2014;40:1328-37.
56. Stefansson H, Meyer-Lindenberg A, Steinberg S, et al. CNVs conferring risk of autism or schizophrenia affect cognition in controls. *Nature* 2014;505:361-6.
57. Giovanoli S, Engler H, Engler A, et al. Stress in puberty unmasks latent neuropathological consequences of prenatal immune activation in mice. *Science* 2013;339:1095-9.
58. Sobin C, Kiley-Brabeck K, Monk SH, et al. Sex differences in the behavior of children with the 22q11 deletion syndrome. *Psychiatry Res* 2009;166:24-34.